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(21) International Application Number: PCT/CA96/00684 (22) International Filing Date: 11 October 1996 (11.10.96) (30) Priority Data: 08/540,753 11 October 1995 (11.10.95) US (60) Parent Application or Grant (63) Related by Continuation US 08/540,753 (CON) Filed on 11 October 1995 (11.10.95) (71) Applicant (for all designated States except US): CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Av- enue West, North York, Ontario M2R 3T4 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): YANG, Yan-Ping [CA/CA]; Apartment 709, 120 Torresdale Avenue, Willow- dale, Ontario M2R 3N7 (CA). MYERS, Lisa, E. [CA/CA]; 187 Elizabeth Street, Guelph, Ontario N1E 2X5 (CA). HARKNESS, Robin, E. [CA/CA]; Apartment #1706, 640 Sheppard Avenue East, Willowdale, Ontario M2K 1B8		(CA). KLEIN, Michel, H. [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA). (74) Agent: STEWART, Michael, I.; Sim & McBurney, 6th floor, 330 University Avenue, Toronto, Ontario M5G 1R7 (US). (81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published With international search report.
(54) Title: TRANSFERRIN RECEPTOR PROTEIN OF MORAXELLA (57) Abstract An isolated and purified non-denatured transferrin receptor protein of a <i>Moraxella</i> strain, particularly <i>M. catarrhalis</i> , has an apparent molecular mass of about 80 to about 90 kDa, as determined by SDS-PAGE. The transferrin receptor protein or a fragment analog thereof is useful in diagnostic applications and immunogenic compositions, particularly for <i>in vivo</i> administration to a host to confer protection against disease caused by a strain of <i>Moraxella</i> .		

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TITLE OF THE INVENTION
TRANSFERRIN RECEPTOR
PROTEIN OF MORAXELLA

FIELD OF THE INVENTION

The present invention relates to the field of immunology and is particularly concerned with transferrin receptor protein from *Moraxella*, methods of production, and uses thereof.

BACKGROUND OF THE INVENTION

Otitis media is the most common illness of early childhood with approximately 80% of all children suffering at least one bout of otitis media before the age of three (ref. 1 - Throughout this application, various references are referred to in parenthesis to more fully describe the state of the art to which this invention pertains. Full bibliographic information for each citation is found at the end of the specification, immediately preceding the claims. The disclosures of these references are hereby incorporated by reference into the present disclosure). Chronic otitis media can lead to hearing, speech and cognitive impairment in children. It is caused by bacterial infection with *Streptococcus pneumoniae* (approximately 50%), non-typable *Haemophilus influenzae* (approximately 30%) and *Moraxella* (*Branhamella*) *catarrhalis* (approximately 20%). In the United States alone, treatment of otitis media costs between one and two billion dollars per year for antibiotics and surgical procedures, such as tonsillectomies, adenoidectomies and insertion of tympanostomy tubes. Because otitis media occurs at a time in life when language skills are developing at a rapid pace, developmental disabilities specifically related to learning and auditory perception have been documented in youngsters with frequent otitis media.

M. catarrhalis mainly colonizes the respiratory tract and is predominantly a mucosal pathogen. Studies

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using cultures of middle ear fluid obtained by tympanocentesis have shown that *M. catarrhalis* causes approximately 20% of cases of otitis media (ref. 2).

Long regarded as an opportunistic pathogen, *B. catarrhalis* is now recognized to cause a variety of potentially debilitating human diseases as a result of localized or, more rarely, systemic infection.

M. catarrhalis is an important cause of lower respiratory tract infections in adults, particularly in the setting of chronic bronchitis and emphysema (refs. 3, 4, 5, 6, 7, 8 and 9). *M. catarrhalis* also causes sinusitis in children and adults (refs. 10, 11, 12, 13 and 14) and occasionally causes invasive disease (refs. 15, 16, 17, 18, 19 and 20). In the hospital setting, *M. catarrhalis* has been suspected in outbreaks of nosocomial infection (ref. 21).

The incidence of otitis media caused by *M. catarrhalis* is increasing. As ways of preventing otitis media caused by pneumococcus and non-typeable *H. influenzae* are developed, the relative importance of *M. catarrhalis* as a cause of otitis media can be expected to further increase. Also antibiotic resistance is becoming common in clinical isolates of *M. catarrhalis* (ref 22). Thus, prior to 1970 no β -lactamase producing *M. catarrhalis* strains had been reported, but since the mid seventies, β -lactamase expressing strains have been detected with ever increasing frequency among isolates. Recent surveys suggest that 75% of clinical isolates produce β -lactamase.

Iron-restriction is a general host defence mechanism against microbial pathogens, however, it is not necessarily growth rate-limiting for all pathogens (ref. 23). A number of bacterial species including *Neisseria meningitidis* (ref. 24), *N. gonorrhoeae* (ref. 25) and *Haemophilus influenzae* (ref. 26), expresses two outer membrane proteins which specifically bind human

transferrin (refs. 27, 28). The expression of these proteins is regulated by the amount of iron in the growth environment. Unlike the receptors of other bacterial species, the *M. catarrhalis* receptors have a preferred
5 affinity for iron-laden (i.e., ferri-) transferrin (ref. 29). The two *M. catarrhalis* transferrin receptors (TfR) have molecular masses of 115 kDa (TfR1) and about 80 to 90 kDa (TfR2) (ref. 27).

The outer membrane protein OMP B2 (refs. 30, 31) has
10 a molecular mass similar to that of TfR2, and it has been reported that the expression of OMP B2 is iron-regulated (ref. 32).

Yu and Schryvers (ref. 29) describe a method for purification of the transferrin receptor proteins TfR1
15 and TfR2 from *M. catarrhalis* by selective elution from a transferrin-sepharose affinity column using the denaturing agent guanidine HCl.

In this method of the preparative isolation of receptor proteins, a suspension of iron-deficient crude
20 *M. catarrhalis* membranes was solubilized by the addition of EDTA to 20 mM and Sarkosyl to 0.75% and then the mixture was centrifuged for 15 minutes at 20,000 x g to remove debris. Twenty ml of Fe₃hTf-Sepharose (i.e. an iron-saturated form of human transferrin) was added to
25 the supernatant and then incubated with mixing at room temperature for 45 minutes. The mixture was applied to a column and after removing the binding solution, the resin was washed with an additional 250 ml of 50 mM Tris-HCl, 1 M NaCl, 20 mM EDTA, 0.75% Sarkosyl, pH 8 buffer
30 containing 250 mM guanidine HCl. The TfR2 protein was eluted using a buffer (lacking Sarkosyl) containing 1.5 M guanidine-HCl and subsequently the TfR1 protein was eluted by the application of a buffer containing 4 M guanidine-HCl. The fractions were dialysed against 50 mM
35 Tris HCl, pH 8 over a 24 hour period.

In a further modification of this procedure, apohTf-Sephacrose was mixed with the solubilized membrane preparation in order to bind TfR1 and then the treated solubilized membrane preparation was exposed to Fe₂hTf-Sephacrose to bind the remaining TfR2. The affinity resins were subsequently washed and the receptor proteins eluted as described above.

M. catarrhalis infection may lead to serious disease. It would be advantageous to provide non-denatured transferrin receptor protein from *M. catarrhalis* for use as antigens in immunogenic preparations including vaccines, carriers for other antigens and immunogens and the generation of diagnostic reagents.

SUMMARY OF THE INVENTION

The present invention is directed towards the provision of purified and isolated transferrin receptor protein of *Moraxella catarrhalis* and other *Moraxella* strains, having an apparent molecular mass of about 80 to 90 kDa.

In accordance with one aspect of the invention, there is provided an isolated and purified, non-denatured transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, as determined by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE), or a fragment or an analog thereof (TfR2). The transferrin receptor protein may be substantially in its native conformation (so as to have substantially retained the characteristic immunogenicity of the transferrin receptor protein in the *Moraxella* strain) and may be isolated from a *M. catarrhalis* strain, such as from *M. catarrhalis* 4223, 5191 or 135. Such isolated and purified transferrin receptor protein of apparent molecular mass of about 80 to about 90 kDa is substantially free from non-80 to 90 kDa *Moraxella* proteins, particularly the OMP B2 protein

and the transferrin receptor protein having an apparent molecular mass of about 105 kDa of the *Moraxella* strains, phospholipids and lipopolysaccharide of *Moraxella*. The about 80 to about 90 kDa transferrin receptor protein is at least about 70 wt% pure, preferably at least about 90 wt% pure, and may be in the form of an aqueous solution thereof.

The present invention also provides an immunogenic composition comprising an immunoeffective amount of an active component, which may be the transferrin receptor protein or fragment or analog thereof, provided herein along with a pharmaceutically acceptable carrier therefor. The immunogenic composition may be formulated as a vaccine for *in vivo* administration to a host to confer protection against diseases caused by a strain of *Moraxella*, particularly *M. catarrhalis*. The immunogenic composition may be formulated as a microparticle capsule, ISCOM or liposome preparation. The immunogenic composition may be used in combination with a targeting molecule for delivery to specific cells of the immune system or to mucosal surfaces. Some targeting molecules include strain B12 and fragments of bacterial toxins, as described in WO 92/17167 (Biotech Australia Pty. Ltd.), and monoclonal antibodies, as described in U.S. Patent No. 5,194,254 (Barber et al). The immunogenic compositions of the invention (including vaccines) may further comprise at least one other immunogenic or immunostimulating material and the immunostimulating material may be at least one adjuvant. Suitable adjuvants for use in the present invention include, (but are not limited to) aluminum phosphate, aluminum hydroxide, QS21, Quil A, derivatives and components thereof, calcium phosphate, calcium hydroxide, zinc hydroxide, a glycolipid analog, an octadecyl ester of an amino acid, a muramyl dipeptide, polyphosphazene and a lipoprotein. Advantageous combinations of adjuvants are

described in copending United States patent Application Serial No. 08/261,194 filed June 16, 1994, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference. The invention further
5 includes an antibody specific for the transferrin receptor protein provided herein producible by immunizing a host with an immunogenic composition as provided herein.

The immunogenic compositions provided herein may be
10 formulated to comprise at least one additional immunogen, which may comprise or be derived from a paramyxovirus, *Chlamydia*, polio, hepatitis B, diphtheria toxoid, tetanus toxoid, influenza, haemophilus, pertussis, pneumococcus, mycobacteria and hepatitis A.

15 In a further aspect of the invention, there is provided a method of generating an immune response in a host, comprising administering thereto an immuno-effective amount of the immunogenic composition as provided herein. The immune response may be a humoral or
20 a cell-mediated immune response. Hosts in which protection against disease may be conferred include primates including humans.

The present invention provides, in an additional aspect thereof, a method of producing a vaccine
25 comprising administering the immunogenic composition provided herein to a test host to determine an amount and a frequency of administration of the transferrin receptor protein to confer protection against disease caused by a strain of *Moraxella* and formulating the transferrin
30 receptor protein in a form suitable for administration to a treated host in accordance with said determined amount and frequency of administration. The treated host may be a human.

A further aspect of the invention provides a method
35 of determining the presence in a sample of antibodies specifically reactive with a transferrin receptor protein

of a strain of *Moraxella* having a molecular mass of about 80 to about 90 kDa, comprising the steps of:

- 5 (a) contacting the sample with the transferrin receptor protein as provided herein to produce complexes comprising the transferrin receptor protein and any said antibodies present in the sample specifically reactive therewith; and
- (b) determining production of the complexes.

10 In a further aspect of the invention, there is provided a method of determining the presence in a sample of a transferrin receptor protein of a strain of *Moraxella* having a molecular mass of about 80 to about 90 kDa, in a sample comprising the steps of:

- 15 (a) immunizing a subject with the immunogenic composition as provided herein, to produce antibodies specific for the transferrin receptor protein;
- (b) contacting the sample with the antibodies to produce complexes comprising any outer membrane protein present in the sample and said outer membrane protein specific antibodies; and
- 20 (c) determining production of the complexes.

25 The transferrin receptor protein may be part of a *Moraxella catarrhalis* strain.

A further aspect of the invention provides a diagnostic kit for determining the presence of antibodies in a sample specifically reactive with the transferrin receptor protein of a strain of *Moraxella* having a molecular mass of about 80 to about 90 kDa, comprising:

- 30 (a) the transferrin receptor protein as provided herein;
- (b) means for contacting the transferrin receptor protein with the sample to produce complexes comprising the transferrin receptor protein
- 35

and any said antibodies present in the sample;
and

- (c) means for determining production of the complexes.

5 The invention also provides a diagnostic kit for detecting the presence, in a sample, of a transferrin receptor protein of a strain of *Moraxella* having a molecular mass of about 80 to about 90 kDa comprising:

- (a) an antibody specific for the about 80 to about
10 90 kDa transferrin receptor protein as provided herein;
- (b) means for contacting the antibody with the sample to produce a complex comprising the transferrin receptor protein and transferrin
15 receptor protein-specific antibody; and
- (c) means for determining production of the complex.

The present invention provides, in a further aspect, a method of producing a vaccine, comprising administering
20 the immunogenic composition provided herein to a test host to determine an amount and a frequency of administration thereof to confer protection against disease caused by a *Moraxella* strain that produces a transferrin receptor protein having an apparent molecular
25 mass of about 80 to about 90 kDa, as determined by SDS-PAGE or a protein capable of inducing antibodies in a test host specifically reactive with the transferrin protein; and formulating the immunogenic composition in a form suitable for administration to a treated host,
30 including a human host, in accordance with the determined amount and frequency of administration.

In an additional aspect of the invention, there is provided a method of producing monoclonal antibodies specific for a transferrin receptor protein of a
35 *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, comprising:

(a) administering the transferrin receptor protein provided herein to at least one mouse to produce at least one immunized mouse,

(b) removing B-lymphocytes from the at least one
5 immunized mouse;

(c) fusing the B-lymphocytes from the at least one immunized mouse with myeloma cells, thereby producing hybridomas;

(d) cloning the hybridomas;

10 (e) selecting clones which produce anti-transferrin receptor protein antibody;

(f) culturing the anti-transferrin receptor protein antibody-producing clones; and then

(g) isolating anti-transferrin receptor protein
15 antibodies from the cultures.

In a further aspect of the invention, there is provided a method of producing an isolated and purified non-denatured transferrin receptor protein of a strain of *Moraxella*, such as *M. catarrhalis* having a molecular mass
20 of about 80 to about 90 kDa, comprising the steps of:

(a) providing a cell mass of the *Moraxella* strain;

(b) selectively extracting aqueous soluble proteins from the cell mass to provide a first supernatant and a first pellet;

25 (c) separating the first supernatant from the first pellet;

(d) selectively solubilizing at least a transferrin receptor protein having a molecular mass of about 80 to 90 kDa from the first pellet to provide a second supernatant and a second pellet;
30

(e) separating the second supernatant from the second pellet; and

(f) purifying transferrin receptor protein having a molecular mass of about 80 to about 90 kDa in the
35 second supernatant substantially free from other

Moraxella proteins solubilized from the first pellet in the selective solubilization step.

The aqueous soluble proteins may be selectively extracted from the cell mass by contacting the cell mass with a buffered aqueous solution, which may comprise about 50 mM to about 1 M Tris-HCl at a pH of about 7 to about 8.5, sonication of the cell mass to disrupt the same, and centrifugation to form the first pellet and the first supernatant. The selective solubilization step may be effected by contacting the first pellet at least once with a buffered aqueous solution, which may be about 20 mM to about 1 M Tris-HCl, comprising a detergent, which may be about 0.2 to about 2 wt.% of TRITON X-100 (Trademark for non-ionic detergent which is octadecylphenol (ethylene glycol) 10), and a solubilizing agent, which may be about 2 to about 20 mM of EDTA, sonication of the first pellet to disrupt the same and centrifugation to form the second pellet and the second supernatant. Such contacting, sonication and centrifugation steps may be effected at least twice and supernatant from each such step is then pooled to provide the second supernatant.

The purification step may be effected by multiple chromatographic column operations, which may include:

- (a) a first chromatographic operation on a first chromatography column, which may be a DEAE-Sephacel column through which the transferrin receptor selectively flows and on which contaminating proteins bind, such as by use of a buffer comprising about 10 mM to about 1 M Tris-HCl at a pH of about 7 to about 8.5,
- (b) a second chromatographic operation on a second chromatography column comprising a cation-exchange matrix, such as SE-Cellulose/D529, through which the transferrin receptor selectively flows and on which contaminating proteins bind, such as by use of a

buffer comprising about 10 mM to about 1 M Tris-HCl at a pH of about 7 to about 8.5,

(c) a third chromatographic operation on a third chromatography column, such as hydroxyapatite, on which the transferrin receptor protein is selectively bound in preference to OMP B2 protein, such as by loading the column at a pH of about 7 to about 8.5, and

(d) eluting the transferrin receptor protein from the third chromatography column, such as by using a buffer solution having a pH of about 7 to about 8.5 containing about 100 to about 250 mM KH_2PO_4 .

The preparation procedure provided herein results in the isolation of the transferrin receptor protein in a purified form and in a nondenatured form.

In this application, the term "a transferrin receptor protein (Tfr2) of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa" is used to define a family of transferrin receptor proteins of *M. catarrhalis* having molecular mass of between about 80 and about 90 kDa and includes proteins having variations in their amino acid sequences including those naturally occurring in various strains of *Moraxella*. In this application, a first protein is a "functional analog" of a second protein if the first protein is immunologically related to and/or has the same function as the second protein. The functional analog may be, for example, a fragment of the protein or a substitution, addition or deletion mutant thereof.

Advantages of the present invention include:

- a method for isolating purified transferrin receptor protein having an apparent molecular mass of about 80 to about 90 kDa from a *Moraxella* strain that produces the transferrin receptor protein, including *Moraxella catarrhalis*;

- an isolated and purified non-denatured transferrin receptor protein having an apparent molecular mass of about 80 to about 90 kDa and isolatable from a *Moraxella* strain; and
- 5 - diagnostic kits and immunological reagents for specific identification of *Moraxella* and hosts infected thereby.

BRIEF DESCRIPTION OF THE FIGURES

The present invention will be further understood from the following description with reference to the
10 Figures, in which:

Figure 1 is a flow diagram of a method for purifying transferrin receptor protein from *Moraxella catarrhalis*, according to one embodiment of the invention;

Figure 2 shows an analysis by SDS-PAGE of isolated
15 and purified transferrin receptor protein having an apparent molecular mass of about 80 to about 90 kDa;

Figure 3 shows the ability of purified transferrin receptor to bind human transferrin;

Figures 4(a), 4(b) and 4(c) show an immunoblot
20 analysis of purified transferrin receptor protein; and

Figures 5(a) and 5(b) are immunoblots to demonstrate the ability of guinea pig anti-Tfr antisera produced by immunization with isolated and purified transferrin receptor protein to specifically distinguish between *M.*
25 *catarrhalis* and other bacterial pathogens that cause otitis media.

GENERAL DESCRIPTION OF INVENTION

The present invention provides novel techniques which can be employed for preparing purified transferrin
30 receptor protein Tfr1 from *M. Catarrhalis*. Any *M. Catarrhalis* strain that produces Tfr2 may be conveniently used to provide the isolated and purified Tfr2 as provided herein. Such strains are generally available from clinical sources and from bacterial culture
35 collections. Appropriate strains of *M. catarrhalis* may include *M. catarrhalis* 5191, 135 and 4223.

Referring to Figure 1, there is illustrated a flow diagram of a method for purifying a transferrin protein from *M. catarrhalis* having an apparent molecular mass of about 80 to about 90 kDa, as determined by SDS-PAGE according to one embodiment of the invention. The various operations described below may be effected at ambient temperatures, generally about 4° to about 30°C. Whole cells are contacted with an aqueous buffer medium, which may contain about 10 mM to about 1 M Tris-HCl at pH about 7 to about 8.5 and disrupted by sonication to produce, following centrifugation, a first supernatant (S1) and a first pellet (PPT1). The supernatant (S1) contains a substantial proportion of the soluble protein from the cell pellet and is discarded. The remaining pellet (PPT1) is selectively detergent extracted to remove any residual soluble proteins and to solubilize membrane proteins, including the TfR2 protein, from the pellet. Such selective extraction may be effected in any convenient manner. One such procedure may involve multiple detergent extractions of the pellet. In particular, a first detergent extraction may be effected using Triton X-100 and EDTA followed by centrifugation to form a second pellet (PPT2) and a second supernatant (S2), which is retained. Following separation of the second supernatant (S2), the residual pellet PPT2 is again detergent extracted using Triton X-100 and EDTA, followed by centrifugation. The resulting third supernatant (S3), following separation from the residual third pellet (PPT3), which is discarded, is combined with the second supernatant (S2) from the first extraction to provide pooled supernatants (TfR2-1). Such Triton X-100 extractions may be effected using a solution of concentration about 0.2 to about 2 wt% Triton X-100 and about 2 to about 20 mM EDTA under buffer conditions, such as a pH about 7 to about 8.5, using about 10 mM to about 1 M Tris.

The pooled supernatants (TfR2-1) contain the TfR2 protein, as well as contaminating *M. catarrhalis* proteins, including the OMP B2 protein which has the same approximate molecular mass (see Lanes 3 and 5, Figure 2).

5 The pooled supernatants (TfR2-1) are processed to remove such impurities in a series of column chromatography operations.

In a first such column chromatography operation, the pooled supernatants (TfR2-1) may be
10 applied to a DEAE-Sephacel column or other suitable column, suitably buffered, such as with about 10 mM to about 1 M Tris-HCl at a pH of about 7 to about 8.5, to permit the TfR2 protein to pass through the column, while impurities are bound to the column. The DEAE-Sephacel
15 column may be washed before further processing of the flow through.

The flow through and wash from the DEAE-Sephacel column (TfR2-2) then may be applied to a cation-exchange column, for example, SE-cellulose/D529 or S-Sepharose,
20 suitably buffered, such as with about 10 mM to about 1 M Tris-HCl at pH about 7 to about 8.5, to permit TfR2 protein to pass through the column, while impurities are bound to the columns. The cation-exchange column may be washed before further processing of the flow-through (see
25 lane 4, Figure 2).

The flow-through and wash from the cation-exchange column (TfR2-3), which still contains the OMP B2 protein, as well as small quantities of residual proteinaceous contaminants, then may be applied to a hydroxyapatite
30 column under such buffer conditions that the TfR2 protein binds to the column in preference to the OMP B2 protein and other contaminating proteins, which pass through the column. Such conditions may be provided using about 5 to about 50 mM potassium phosphate buffer at pH about 6 to
35 about 8.5. The flow-through contains the contaminating proteins (see lane 5, Figure 2).

Following repeated washing with the buffer to remove column-retained contaminants, the TfR2 protein then is eluted from the hydroxyapatite column using a suitable buffer, such as about 150 to about 250 mM potassium phosphate buffer solution at pH about 7 to about 8.5. The eluted fractions are collected, aliquots analyzed by SDS-PAGE and TfR2-containing fractions pooled. The pooled fractions may be concentrated to provide a TfR2 protein-containing solution (see lane 6, Figure 2). By this procedure, there is provided an aqueous solution of an isolated and purified non-denatured TfR2 protein. The aqueous TfR2 solution may be provided in any convenient concentration consistent with the intended use of the material, generally about 100 to about 200 µg/ml.

Referring to Figure 2, there is shown an analysis of the purity of transferrin receptor protein by SDS-PAGE analysis, purified by the method described herein as typified by that schematically shown in Figure 1. In Figure 2, Lane 1 shows the *M. catarrhalis* cell lysate. Lane 2 shows the pooled supernatants (TfR2-1). Lane 3 shows the run-through fraction following DEAE-Sephacel column chromatography (TfR2-2) and Lane 4 shows the run-through fraction of SE-cellulose/D529 column chromatography (TfR2-3). Lane 5 shows the flow-through fraction of HTP and contains the OMP B2 protein. Lane 6 shows the HTP bound fraction containing TfR2. Lane 7 shows transferrin receptor protein TfR2 isolated by affinity chromatography essentially as described in ref. 29. The method for purification of TfR2 protein as provided herein produces at least a 70% pure TfR2 protein preparation. The illustrated preparation in Lane 6 in Figure 2 is at least 95% pure, as determined by densitometry scanning.

Referring to Figure 3, there is illustrated the ability of purified TfR2 of *M. catarrhalis* as provided herein to specifically bind human transferrin. Proteins

were separated by SDS-PAGE and in vitro transferrin binding assessed essentially as described in ref. 27. Lane 1 contains TfR2 of *M. catarrhalis* isolated by affinity chromatography essentially as described in ref. 29. Lane 2 contains TfR2 from *M. catarrhalis* purified as described herein. Lane 3 contains *M. catarrhalis* OMPB2 protein. As can be seen from Figure 3, the transferrin binding protein preparations specifically bind transferrin. These results confirm the isolation of TfR2 from *M. catarrhalis* by the method provided herein.

To be useful as a component of immunogenic compositions (including vaccines) and as an antigen in diagnostic embodiments, the transferrin receptor protein purified as described herein should advantageously be capable of generating antibodies that recognize or neutralize *M. catarrhalis* strains, including a plurality thereof.

Referring to Figures 4(a), 4(b) and 4(c), there is shown an immunoblot analysis of TfR2 as provided herein and shows the specific separation of TfR2 from OMP B2 protein. In each of the Figures, sample 1 contains OMP B2 protein and sample 2 contains TfR2 protein. The *M. catarrhalis* strains were a: strain 5191; b: strain 135; and c: strain 4223. Samples were separated by SDS-PAGE. Sample 3 was purified TfR2. Figure 4(a) shows a Coomassie blue stained gel. Figure 4(b) shows an immunoblot using an anti-TfR2 antiserum. Figure 4(c) shows an immunoblot using an anti-OMP B2 antiserum. The results of Figure 4 confirm the specific purification of transferrin receptor protein TfR2 from *M. catarrhalis* by the method provided herein and particularly substantially free from OMBP2 protein.

Referring to Figure 5(b), there is illustrated an immunoblot showing the ability of guinea pigs anti-TfR antisera produced by immunizing guinea pigs with purified TfR protein as provided herein to recognize TfR2 protein

from *M. catarrhalis* isolated from a variety of sources. The samples tested were as follows:

<u>Lane</u>	<u><i>M. catarrhalis</i></u>	<u>Source</u>
1	Purified Tfr2	<i>M. catarrhalis</i> 4223
5 2	56	Middle ear fluid
3	4223	Middle ear fluid

In one embodiment of the present invention, the isolated and purified Tfr2 protein as provided herein is useful for generating antibodies that can be used to specifically distinguish *M. catarrhalis* from other bacterial pathogens that cause otitis media. Thus referring to Figures 5(a) and 5(b), there is illustrated an SDS-PAGE and an immunoblot respectively showing the specific reactivity of guinea pig anti-Tfr2 antisera produced by immunizing guinea pigs with Tfr2 protein as provided herein. The samples analyzed were as follows:

<u>Lane</u>	<u>Bacterium</u>	<u>Source</u>
1	Tfr2 protein	<i>M. catarrhalis</i> 4223
2	<i>M. catarrhalis</i> 56	Middle ear fluid
20 3	<i>M. catarrhalis</i> 4223	Middle ear fluid
4	<i>H. influenzae</i> strain 12	
5	<i>H. influenzae</i> strain 30	Otitis media isolate
6	<i>H. influenzae</i> LCOC 1	Otitis media isolate
7	<i>S. pneumoniae</i>	ATCC 6303
25 8	<i>S. pneumoniae</i>	ATCC 6304
9	<i>S. pneumoniae</i>	ATCC 6314

The results shown in Figure 5(b) clearly show the usefulness of Tfr2-specific antisera as provided herein to distinguish between bacterial pathogens that produce diseases with similar clinical symptoms.

Results shown in Table 1 below illustrate the ability of guinea pig anti-Tfr2 antisera produced by immunization with Tfr2 protein as provided herein, to kill *M. catarrhalis*. The results show that antisera

produced by immunization with Tfr2 protein isolated from strain 4223 were bactericidal against a homologous non-clumping *M. catarrhalis* strain RH408 derived from strain 4223. The ability of the isolated and purified Tfr2 protein as provided herein to generate bactericidal antibodies is *in vivo* evidence of utility of the Tfr2 protein as provided herein as a vaccine to protect against diseases caused by *M. catarrhalis*.

Thus, in accordance with another aspect of the present invention, there is provided a vaccine against *Moraxella* or other bacterial pathogens that produce Tfr2 protein or produce a protein capable of inducing antibodies that specifically recognize Tfr2, comprising an immunogenically-effective amount of Tfr2 protein as provided herein and a physiologically-acceptable carrier therefor. The Tfr2 protein provided herein also may be used as a carrier protein for hapten, polysaccharides or peptides to make a conjugate vaccine against antigenic determinants unrelated to Tfr2.

The Tfr2 protein provided herein is useful as a diagnostic reagent, as an antigen or for the generation of anti-Tfr2 antibodies, antigen for vaccination against the diseases caused by species of *Moraxella*.

In additional embodiments of the present invention, the Tfr2 protein as provided herein may be used as a carrier molecule to prepare chimeric molecules and conjugate vaccines (including glycoconjugates) against pathogenic bacteria, including encapsulated bacteria. Thus, for example, glycoconjugates of the present invention may be used to confer protection against disease and infection caused by any bacteria having polysaccharide antigens including lipooligosaccharides (LOS) and PRP. Such bacterial pathogens may include, for example, *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Escherichia coli*, *Neisseria meningitidis*, *Salmonella typhi*, *Streptococcus mutants*, *Cryptococcus*

neoformans, *Klebsiella*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Particular antigens which can be conjugated to TfR2 protein and methods to achieve such conjugations are described in published PCT application
5 WO 94/12641, assigned to the assignee hereof and the disclosure of which is hereby incorporated by reference thereto.

In another embodiment, the carrier function of TfR2 protein may be used, for example, to induce immunity
10 toward abnormal polysaccharides of tumor cells, or to produce anti-tumor antibodies that can be conjugated to chemotherapeutic or bioactive agents.

The present invention extends to the TfR2 protein for use as a pharmaceutical substance, particularly as an
15 active ingredient in a vaccine against diseases caused by infection of a strain of *Moraxella*.

In a further aspect, the invention provides the use of the TfR2 protein for the preparation of a medicament for immunization against diseases caused by infection
20 with a strain of *Moraxella*.

It is clearly apparent to one skilled in the art, that the various embodiments of the present invention have many applications in the fields of vaccination, diagnosis, treatment of *Moraxella* infections and the
25 generation of immunological reagents. A further non-limiting discussion of such uses is further presented below.

1. Vaccine Preparation and Use

Immunogenic compositions, suitable to be used as
30 vaccines, may be prepared from immunogenic transferrin receptor, proteins or analogs as disclosed herein. Preferably, the antigenic material is extensively dialyzed to remove undesired small molecular weight molecules and/or myophilized for more ready formulation
35 into a desired vehicle. The immunogenic composition elicits an immune response which produces antibodies,

including anti-transferrin receptor antibodies and antibodies that are opsonizing or bactericidal. Should the vaccinated subject be challenged by *Moraxella* or other bacteria that produce a transferrin receptor, the
5 antibodies bind to the transferrin receptor and thereby prevent access of the bacteria to an iron source which is required for viability. Furthermore, opsonizing or bactericidal anti-TfR antibodies may also provide protection by alternative mechanisms.

10 Immunogenic compositions including vaccines may be prepared as injectables, as liquid solutions or emulsions. The transferrin receptor protein may be mixed with pharmaceutically acceptable excipients which are compatible therewith. Such excipients may include,
15 water, saline, dextrose, glycerol, ethanol, and combinations thereof. The immunogenic compositions and vaccines may further contain auxiliary substances, such as wetting or emulsifying agents, pH buffering agents, or adjuvants to enhance the effectiveness thereof.
20 Immunogenic compositions and vaccines may be administered parenterally, by injection subcutaneously or intramuscularly. Alternatively, the immunogenic compositions formed according to the present invention, may be formulated and delivered in a manner to evoke an
25 immune response at mucosal surfaces. Thus, the immunogenic composition may be administered to mucosal surfaces by, for example, the nasal or oral (intragastric) routes. Alternatively, other modes of administration including suppositories and oral
30 formulations may be desirable. For suppositories, binders and carriers may include, for example, polyalkalene glycols or triglycerides. Such suppositories may be formed from mixtures containing the active ingredient(s) in the range of about 0.5 to about
35 10%, preferably about 1 to 2%. Oral formulations may include normally employed incipients such as, for

example, pharmaceutical grades of saccharine, cellulose and magnesium carbonate. These compositions can take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain
5 about 1 to 95% of the transferrin receptor protein, preferably about 20 to about 75%.

The immunogenic preparations and vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be
10 therapeutically effective, protective and immunogenic. The quantity to be administered depends on the subject to be treated, including, for example, the capacity of the individual's immune system to synthesize antibodies, the degree of protection desired and if needed, to produce a
15 cell-mediated immune response. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner. However, suitable dosage ranges are readily determinable by one skilled in the art and may be of the order of micrograms of the transferrin
20 receptor protein per vaccination. Suitable regimes for initial administration and booster doses are also variable, but may include an initial administration followed by subsequent administrations. The dosage may also depend on the route of administration and will vary
25 according to the size of the host.

The concentration of the transferrin receptor protein in an immunogenic composition according to the invention is in general about 1 to 95%. A vaccine which contains antigenic material of only one pathogen is a
30 monovalent vaccine. Vaccines which contain antigenic material of several pathogens are combined vaccines and also belong to the present invention. Such combined vaccines contain, for example, material from various pathogens or from various strains of the same pathogen,
35 or from combinations of various pathogens.

Immunogenicity can be significantly improved if the antigens are co-administered with adjuvants, commonly used as 0.05 to 0.1 percent solution in phosphate-buffered saline. Adjuvants enhance the immunogenicity of an antigen but are not necessarily immunogenic themselves. Adjuvants may act by retaining the antigen locally near the site of administration to produce a depot effect facilitating a slow, sustained release of antigen to cells of the immune system. Adjuvants can also attract cells of the immune system to an antigen depot and stimulate such cells to elicit immune responses.

Immunostimulatory agents or adjuvants have been used for many years to improve the host immune responses to, for example, vaccines. Intrinsic adjuvants, such as lipopolysaccharides, normally are the components of the killed or attenuated bacteria used as vaccines. Extrinsic adjuvants are immunomodulators which are typically non-covalently linked to antigens and are formulated to enhance the host immune responses. Thus, adjuvants have been identified that enhance the immune response to antigens delivered parenterally. Some of these adjuvants are toxic, however, and can cause undesirable side-effects, making them unsuitable for use in humans and many animals. Indeed, only aluminum hydroxide and aluminum phosphate (collectively commonly referred to as alum) are routinely used as adjuvants in human and veterinary vaccines. The efficacy of alum in increasing antibody responses to diphtheria and tetanus toxoids is well established and a HBsAg vaccine has been adjuvanted with alum. While the usefulness of alum is well established for some applications, it has limitations. For example, alum is ineffective for influenza vaccination and inconsistently elicits a cell mediated immune response. The antibodies elicited by alum-adjuvanted antigens are mainly of the IgG1 isotype

in the mouse, which may not be optimal for protection by some vaccinal agents.

A wide range of extrinsic adjuvants can provoke potent immune responses to antigens. These include
5 saponins complexed to membrane protein antigens (immune stimulating complexes), pluronic polymers with mineral oil, killed mycobacteria in mineral oil, Freund's complete adjuvant, bacterial products, such as muramyl dipeptide (MDP) and lipopolysaccharide (LPS), as well as
10 lipid A, and liposomes.

To efficiently induce humoral immune responses (HIR) and cell-mediated immunity (CMI), immunogens are often emulsified in adjuvants. Many adjuvants are toxic, inducing granulomas, acute and chronic inflammations
15 (Freund's complete adjuvant, FCA), cytolysis (saponins and Pluronic polymers) and pyrogenicity, arthritis and anterior uveitis (LPS and MDP). Although FCA is an excellent adjuvant and widely used in research, it is not licensed for use in human or veterinary vaccines because
20 of its toxicity.

Desirable characteristics of ideal adjuvants include:

- (1) lack of toxicity;
- (2) ability to stimulate a long-lasting immune response;
- 25 (3) simplicity of manufacture and stability in long-term storage;
- (4) ability to elicit both CMI and HIR to antigens administered by various routes, if required;
- (5) synergy with other adjuvants;
- 30 (6) capability of selectively interacting with populations of antigen presenting cells (APC);
- (7) ability to specifically elicit appropriate T_H1 or T_H2 cell-specific immune responses; and
- (8) ability to selectively increase appropriate antibody
35 isotype levels (for example, IgA) against antigens.

U.S. Patent No. 4,855,283 granted to Lockhoff et al on August 8, 1989 which is incorporated herein by reference thereto teaches glycolipid analogues including N-glycosylamides, N-glycosylureas and N-glycosylcarbamates, each of which is substituted in the sugar residue by an amino acid, as immuno-modulators or adjuvants. Thus, Lockhoff et al. (U.S. Patent No. 4,855,283 and ref. 33) reported that N-glycolipid analogs displaying structural similarities to the naturally-occurring glycolipids, such as glycosphingolipids and glycoglycerolipids, are capable of eliciting strong immune responses in both herpes simplex virus vaccine and pseudorabies virus vaccine. Some glycolipids have been synthesized from long chain-alkylamines and fatty acids that are linked directly with the sugars through the anomeric carbon atom, to mimic the functions of the naturally occurring lipid residues.

U.S. Patent No. 4,258,029 granted to Moloney, assigned to the assignee hereof and incorporated herein by reference thereto, teaches that octadecyl tyrosine hydrochloride (OTH) functioned as an adjuvant when complexed with tetanus toxoid and formalin inactivated type I, II and III poliomyelitis virus vaccine. Also, Nixon-George et al. (ref. 34), reported that octadecyl esters of aromatic amino acids complexed with a recombinant hepatitis B surface antigen, enhanced the host immune responses against hepatitis B virus.

2. Immunoassays

The transferrin receptor protein of the present invention is useful as an immunogen for the generation of anti-transferrin receptor protein antibodies, as an antigen in immunoassays including enzyme-linked immunosorbent assays (ELISA), RIAs and other non-enzyme linked antibody binding assays or procedures known in the art for the detection of anti-bacterial, anti-Moraxella, and anti-TfR antibodies. In ELISA assays, the

transferrin receptor protein is immobilized onto a selected surface, for example, a surface capable of binding proteins such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed transferrin receptor protein, a nonspecific protein such as a solution of bovine serum albumin (BSA) that is known to be antigenically neutral with regard to the test sample may be bound to the selected surface. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific bindings of antisera onto the surface.

The immobilizing surface is then contacted with a sample, such as clinical or biological materials, to be tested in a manner conducive to immune complex (antigen/antibody) formation. This may include diluting the sample with diluents, such as solutions of BSA, bovine gamma globulin (BGG) and/or phosphate buffered saline (PBS)/Tween. The sample is then allowed to incubate for from 2 to 4 hours, at temperatures such as of the order of about 25° to 37°C. Following incubation, the sample-contacted surface is washed to remove non-immunocomplexed material. The washing procedure may include washing with a solution, such as PBS/Tween or a borate buffer. Following formation of specific immunocomplexes between the test sample and the bound transferrin receptor protein, and subsequent washing, the occurrence, and even amount, of immunocomplex formation may be determined by subjecting the immunocomplex to a second antibody having specificity for the first antibody. If the test sample is of human origin, the second antibody is an antibody having specificity for human immunoglobulins and in general IgG. To provide detecting means, the second antibody may have an associated activity such as an enzymatic activity that will generate, for example, a colour development upon

incubating with an appropriate chromogenic substrate. Quantification may then be achieved by measuring the degree of colour generation using, for example, a spectrophotometer.

5

EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific Examples. These Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations.

10

Methods of molecular genetics, protein biochemistry, and immunology used but not explicitly described in this disclosure and these Examples are amply reported in the scientific literature and are well within the ability of those skilled in the art.

15

Example 1

This Example illustrates the growth of *M. catarrhalis*.

The *M. catarrhalis* strains used were 135, 4223 (ref. 35), 5191 (ref. 35) (all middle ear isolates), ATCC 25240, Q8 (expectorate isolate) and RH408.

25

To provide cells for isolation of Tfr2, *M. catarrhalis* strains were routinely maintained on chocolate agar plates (BBL).

30

M. catarrhalis strain 4223 was inoculated into 20 mL of brain heart infusion (BHI) broth. The culture was incubated overnight with aeration at 37°C. For growth under iron-restricted conditions, one mL of the overnight-culture was inoculated into 20 mL of BHI broth containing 25 μ M EDDA and the culture was grown at 37°C for approximately 3 to 4 h. Cells grown to mid-log phase

35

($A_{578} > 0.5$) were harvested by centrifugation at 10,000 x g for 20 min. The pellet was used for extraction of transferrin receptor (TfR2) protein as described in Example 3 below.

5 Example 2

 This Example illustrates the generation of a non-clumping strain (RH408) of *M. catarrhalis*.

M. catarrhalis strain 4223 was inoculated into several flasks containing 20 mL of BHI broth, and the
10 cultures were incubated with shaking (170 rpm) overnight at 37°C. Five mL of each overnight culture were transferred to individual 1 mL tubes, and were left sitting undisturbed at room temperature for 3 to 8 hours, to allow bacteria to sediment. One hundred µL of the
15 cleared upper phase of each culture medium were used to inoculate 25 mL of BHI broth and cultures were incubated overnight at 37°C, as described above. This passaging was repeated six times, using 25 µL of cleared medium to
20 inoculate 25 mL of BHI for each overnight culture. Non-clumping bacterial cultures were identified by measuring the A_{578} at intervals over a 3 hour time period, in order to compare the sedimentation rates of the passaged strains to that of the original *M. catarrhalis* strain 4223 culture. Non-clumping mutants, including *M.*
25 *catarrhalis* RH408, did not aggregate during the three hour time period. On BHI agar plates, strain RH408 had a colony morphology typical for all non-clumping strains. Strain RH408 is deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD
30 20852, U.S.A., under the terms of the Budapest Treaty, on December 13, 1994 and was given ATCC Accession No. 55637.

Example 3

 This Example illustrates the extraction of
35 transferrin receptor protein TfR2 by affinity purification.

Affinity purified TfR2 was prepared from total membranes as described in reference 29 using Sepharose 4B (Pharmacia) conjugated human transferrin (Sigma). Bacterial receptor proteins bound to the affinity matrix were selectively eluted using a buffer of 50 mM Tris-HCl pH 8.0, 1M NaCl, 10 mM EDTA, 0.05% sodium lauryl sarkosinate, 2M guanidine. Eluted proteins were dialyzed against 50 mM Tris-HCl pH 8.0.

Example 4

10 This Example illustrates the extraction and purification of transferrin receptor protein TfR2.

Transferrin receptor protein was isolated from *M. catarrhalis* by the procedure generally illustrated in Figure 1.

15 A cell pellet from iron-deprived overnight cultures of *M. catarrhalis* was resuspended in 50 mM Tris-HCl, pH 8.0, and disrupted by sonication. The sonicate was centrifuged at 20,000 x g for 30 minutes and the resultant supernatant (S1), which contained soluble
20 proteins, was discarded. The pellet (PPT1) was suspended in 50 mM Tris, pH 8.0 containing 0.5% Triton X-100 and 10 mM EDTA. This extraction step solubilized TfR2. The suspension was centrifuged at 20,000 x g for 20 minutes to remove insoluble material and the supernatant was
25 applied to a DEAE-Sephacel (Pharmacia) column equilibrated in 50 mM Tris-HCl buffer, pH 8.0. TfR2 was recovered in the flow-through fraction and was further purified using a SE-cellulose D529 column equilibrated in
30 50 mM Tris-HCl, pH 8.0. The flow-through fraction containing TfR2 was next loaded onto a hydroxyapatite (HTP) column equilibrated in 10 mM phosphate buffer, pH 8.0. This step separated OMP B2 (present in the HTP flow-through fraction) from TfR2 which remained bound to the HTP. After extensive washing of HTP column with
35 50 mM phosphate buffer, pH 8.0, TfR2 was eluted from the

matrix with a 200 mM phosphate buffer, pH 8.0. The purity of Tfr2 was assessed by SDS-PAGE analysis.

Example 5

This Example describes the purification of *M. catarrhalis* outer membrane protein B2 OMP B2.

The flow-through fraction of HTP column was analyzed by 12.5% SDS-PAGE. Following electrophoresis, a gel slice corresponding to the OMP B2 protein band was cut from the gel. Protein was recovered from the gel slice by electroelution for 12 hours at 100 volts in elution buffer (15mM NH_4CO_3 /0.1% SDS).

Example 6

This Example illustrates the immunization of guinea pigs.

Guinea pigs (Charles River, Quebec) were immunized intramuscularly (i.m.) on day 1 with a 5 μg dose of either Tfr2 or OMP B2 protein emulsified in complete Freund's adjuvant (CFA). Animals were boosted on days +14 and +28 with the same dose of protein emulsified in incomplete Freund's adjuvant (IFA). Blood samples were taken on day +42. The results obtained are shown in Table 1.

Example 7

This Example describes the analysis of purified Tfr2 and OMP B2 proteins.

Proteins were separated by SDS-PAGE as described by Lugtenberg et al (ref. 36) using 11.5% or 12.5% (w/v) acrylamide (BRL) in the separating gels. Proteins were visualized by staining gels with Coomassie brilliant blue (BioRad). Protein standard molecular mass markers were from BioRad and Pharmacia.

Proteins separated by SDS-PAGE were electroblotted by the method of Towbin et al (ref. 37) onto polyvinylidene difluoride membranes (PVDF, Millipore). For immunoblots, antisera were used at dilutions of 1:1,000. Recombinant protein G horseradish peroxidase

conjugate (Zymed) was used as a reporter at a dilution of 1:4,000. Blots were developed using either a colourimetric reaction (4CN/DAB; Pierce) or a chemiluminescent reaction (LumiGlo; Kirkegaard and Perry). Procedures recommended by the manufacturers were followed for both detection methods. Prestained molecular mass markers were purchased from BioRad.

In vitro transferrin-binding activity of the purified proteins was assessed following the method described by Schryvers and Lee (ref. 27) with some modifications. Briefly, protein samples were separated by SDS-PAGE and then electrophoretically transferred to PCDF membranes. The membranes were incubated with horseradish peroxidase conjugated human transferrin (1:50 dilution; Jackson ImmunoResearch) overnight at 4°C. Blots were developed using the LumiGlo chemiluminescent reaction described above.

Example 8

This Example illustrates the bactericidal assay against *B. catarrhalis*.

Samples (25 µL) of antiserum, were heated to 56°C for 30 minutes to remove complement activity and diluted 1:8 in veronal buffer (NaCl 8.0 g/L, NaHCO₃ 0.25 g/L, Na Barbiturate 0.30 g/L, Barbituric acid 0.45 g/L, MgCl₂.6H₂O 0.1 g/L, CaCl₂.2H₂O 0.05 g/L) containing 0.1% BSA (VBS), and then added to the first well of a 96-well Nunc microtiter plate. Two-fold serial dilutions of antisera in VBS were placed in the remaining wells. Bacterial cells grown to an OD₅₇₈ > 0.5 were diluted 1:200,000 in VBS and 25-µL portions of the bacterial suspension were added to each well. A guinea pig complement (Biowhittaker, Walkersville, MD) was diluted 1:10 in VBS and 25 µL of the solution were added to each well to initiate reactions. The plates were incubated at 37°C for 60 min, and 50 µL of each reaction mixture was then plated onto Mueller-Hinton agar plate containing 2.2%

Mueller-Hinton broth and 1.5% agar. After incubation at 37°C for 48 hours, colonies were counted to determine the bactericidal titer (the reciprocal of the highest dilution of antiserum capable of killing greater than 50% of bacteria compared with controls containing pre-immune sera). The results are shown in Table 1.

SUMMARY OF THE DISCLOSURE

In summary of this disclosure, the present invention provides a transferrin receptor protein having an apparent molecular mass of about 80kDa to about 90kDa and isolated from *M. catarrhalis*, methods of making the same and uses thereof as immunogenic compositions and diagnostic embodiments. Modifications are possible within the scope of this invention.

TABLE 1

Anti-TfR2 antibody titres and bactericidal activity of guinea pig antisera obtained by immunization with transferrin receptor protein (TfR2) from *M. catarrhalis* 4223

<u>Immunogen</u>	<u>Bactericidal Titre¹</u>		<u>Anti-TfR2 Titre²</u>	
	<u>Pre-immune</u>	<u>Post-immune</u>	<u>Pre-immune</u>	<u>Post-immune</u>
TfR2	< 3.0	13.5	<0.5	14.0
OMP B2	< 3.0	< 3.0	<0.5	<0.5

1 Bactericidal titres: expressed in log₂ as the dilution of antiserum capable of killing 50% of cells of *M. catarrhalis* 4223.

2 Anti-TfR2 titres: expressed as log₂ (titre/100)

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CLAIMS

What we claim is:

1. An isolated and purified non-denatured transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, as determined by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE), or a fragment, or an analog thereof.
2. The protein of claim 1 wherein the *Moraxella* strain is *Moraxella catarrhalis*.
3. The protein of claim 2 wherein the strain is *Moraxella catarrhalis* 4223, 5191 or 135.
4. The protein of claim 1 which is at least about 70 wt% pure.
5. The protein of claim 4 which is at least about 90 wt% pure.
6. The protein of claim 1 in the form of an aqueous solution thereof.
7. The protein of claim 1 substantially free from the OMP B2 protein of the *Moraxella* strain.
8. The protein of claim 1 substantially free from the transferrin receptor protein of the *Moraxella* strain having an apparent molecular mass of about 105 kDa.
9. An immunogenic composition, comprising an immunoeffective amount of the protein of claim 1.
10. The immunogenic composition of claim 8 formulated as a vaccine for *in vivo* administration to a host to confer protection against disease caused by a strain of *Moraxella*.
11. The immunogenic composition of claim 10 wherein the strain is *Moraxella catarrhalis*.
12. The immunogenic composition of claim 10 formulated as a microparticle, capsule, ISCOM or liposome preparation.
13. The immunogenic composition of claim 10 in combination with a targeting molecule for delivery to

specific cells of the immune system or to mucosal surfaces.

14. The immunogenic composition of claim 10 further comprising a least one other immunogenic or immunostimulating material.

15. The immunogenic composition of claim 10 wherein the at least one other immunostimulating material is at least one adjuvant.

16. The immunogenic composition of claim 13 wherein the at least one adjuvant is selected from the group consisting of aluminum phosphate, aluminum hydroxide, QS21, Quil A, or components or derivatives thereof, calcium phosphate, calcium hydroxide, zinc hydroxide, a glycolipid analog, an octodecyl ester of an amino acid, a muramyl dipeptide, a lipoprotein, polyphosphazene, ISCOM matrix, ISCOPREP, DC-chol and DDBA.

17. The immunogenic composition of claim 16 wherein the host is a primate.

18. The immunogenic composition of claim 17 wherein the primate is a human.

19. An antibody specific for a transferrin receptor protein of *Moraxella* producible by immunizing a host with the immunogenic composition of claim 9.

20. A method of generating an immune response in a host, comprising administering thereto an immuno-effective amount of the immunogenic composition of claim 9.

21. The method of claim 20 wherein the immune response is a humoral or a cell-mediated immune response.

22. A method of determining the presence in a sample of antibodies specifically reactive with a transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, comprising the steps of:

- (a) contacting the sample with the protein of claim 1 to produce complexes comprising the protein and

any said antibodies present in the sample specifically reactive therewith; and

(b) determining production of the complexes.

23. A method of determining the presence in a sample of a transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, comprising the steps of:

(a) immunizing a subject with the immunogenic composition of claim 9 to produce antibodies specific for the transferrin receptor protein;

(b) contacting the sample with the antibodies to produce complexes comprising any transferrin receptor protein present in the sample and said transferrin receptor protein specific antibodies; and

(c) determining production of the complexes.

24. A diagnostic kit for determining the presence of antibodies in a sample specifically reactive with a transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, comprising:

(a) the transferrin receptor protein of claim 1;

(b) means for contacting the transferrin receptor protein with the sample to produce complexes comprising the transferrin receptor protein and any said antibodies present in the sample; and

(c) means for determining production of the complexes.

25. A diagnostic kit for detecting the presence in a sample of a transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa protein, comprising:

(a) an antibody of claim 19;

(b) means for contacting the antibody with the sample to produce a complex comprising transferrin

receptor protein and transferrin receptor protein-specific antibody; and

(c) means for determining production of the complex.

26. A method for producing a vaccine, comprising:

administering the immunogenic composition of claim 9 to a test host to determine an amount and a frequency of administration thereof to confer protection against disease caused by a *Moraxella* strain that produces a transferrin receptor protein having an apparent molecular mass of about 80 to about 90 kDa, as determined by SDS-PAGE or a protein capable of inducing antibodies in a test host specifically reactive with the transferrin receptor protein; and

formulating the immunogenic composition in a form suitable for administration to a treated host in accordance with said determined amount and frequency of administration.

27. The method of claim 26 wherein the treated host is a human.

28. A method of producing monoclonal antibodies specific for a transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, as determined by SDS-PAGE, comprising:

(a) administering the transferrin receptor protein of claim 1 to at least one mouse to produce at least one immunized mouse;

(b) removing B-lymphocytes from the at least one immunized mouse;

(c) fusing the B-lymphocytes from the at least one immunized mouse with myeloma cells, thereby producing hybridomas;

(d) cloning the hybridomas;

(e) selecting clones which produce anti-transferrin receptor protein antibody;

(f) culturing the anti-transferrin receptor protein antibody-producing clones; and then

(g) isolating anti-transferrin receptor protein antibodies from the cultures.

29. A method of producing an isolated and purified non-denatured transferrin receptor protein of a *Moraxella* strain having an apparent molecular mass of about 80 to about 90 kDa, comprising the steps of:

- (a) providing a cell mass of the *Moraxella* strain;
- (b) selectively extracting aqueous soluble proteins from said cell mass to provide a first supernatant and a first pellet;
- (c) separating said first supernatant from said first pellet;
- (d) selectively solubilizing at least a transferrin receptor protein having a molecular mass of about 80 to 90 kDa from said first pellet to provide a second supernatant and a second pellet;
- (e) separating said second supernatant from said second pellet; and
- (f) purifying transferrin receptor protein having a molecular mass of about 80 to 90 kDa in said second supernatant substantially free from other *Moraxella* proteins solubilized from said first pellet in said selective solubilization step.

30. The method of claim 29 wherein said aqueous soluble proteins are selectively extracted from said cell mass by contacting the cell mass with a buffered aqueous solution, sonicating said cell mass to disrupt the same, and centrifuging to form said first pellet and said first supernatant.

31. The method of claim 30 wherein said buffered aqueous solution comprises about 50 mM to about 1M of Tris-HCl at a pH of about 7 to about 8.5.

32. The method of claim 29 wherein said selective solubilization step is effected by contacting the first

pellet at least once with a buffered aqueous solution comprising a detergent and a solubilizing agent, sonicating of said first pellet to disrupt the same, and centrifuging to form said second pellet and said second supernatant.

33. The method of claim 32 wherein said buffered aqueous solution has a pH of about 7 to about 8.5 and comprises about 10 mM to about 1 M of Tris-HCl, about 0.2 to about 2 wt.% of detergent and about 2 to about 20 mM of EDTA.

34. The method of claim 33 wherein said detergent is Triton X-100.

35. The method of claim 32 wherein said contacting, sonicating and centrifuging steps are effected at least twice and supernatant from each such steps is pooled to provide said second supernatant.

36. The method of claim 29 wherein said purification step is effected by multiple chromatographic column operations.

37. The method of claim 36 wherein said multiple chromatographic column operations includes:

- (a) a first chromatographic operation on a first chromatography column through which said transferrin receptor protein selectively flows and on which contaminating proteins bind;

- (b) a second chromatographic operation on a second chromatography column comprising a cation-exchange matrix through which said transferrin receptor protein selectively flows and on which contaminating proteins bind;

- (c) a third chromatographic operation on a third chromatography column on which said transferrin receptor protein is selectively bound in preference to OMP B2 protein; and

- (d) eluting said transferrin receptor protein from said third chromatography column.

38. The method of claim 37 wherein said first chromatographic operation is effected using about 10 mM to about 1 M Tris-HCl and a pH of about 7 to about 8.5 on a DEAE-Sephacel column.

39. The method of claim 38 wherein said second chromatographic operation is effected using about 10 mM to about 1 M Tris-HCl at a pH of about 7 to about 8.5 on a SE-cellulose column.

40. The method of claim 38 wherein said third chromatographic operation is effected by loading said third chromatographic column which is hydroxyapatite using about 5 to about 50 mM of KH_2PO_4 buffer solution at a pH about 7 to about 8.5, and said elution step is effected by eluting the loaded column using a buffer solution having a pH of about 7 to about 8.5 containing about 150 to about 250 mM KH_2PO_4 .

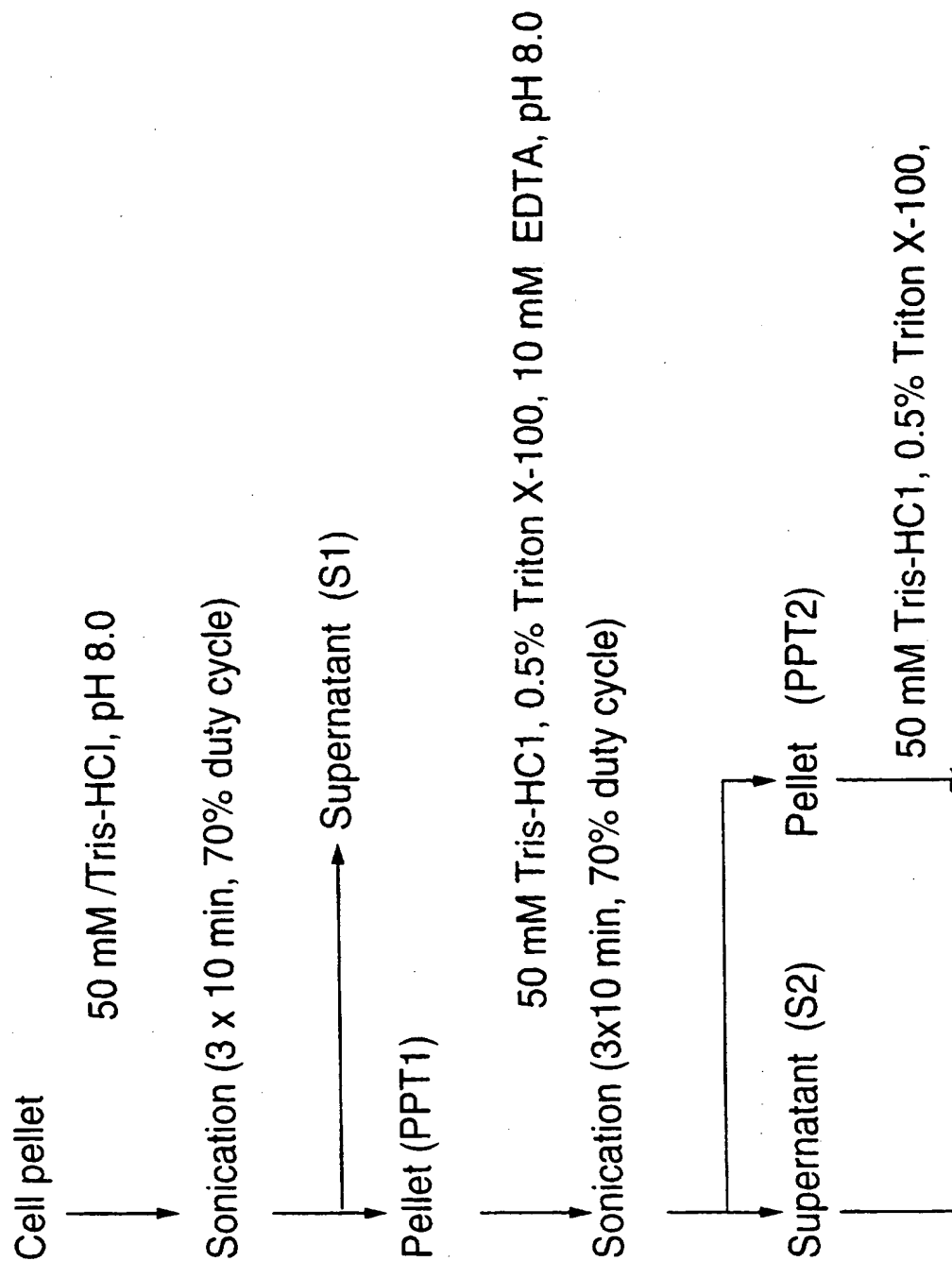
Purification of the outermembrane protein TFR2 from *B. catarrhalis*

FIG.1A

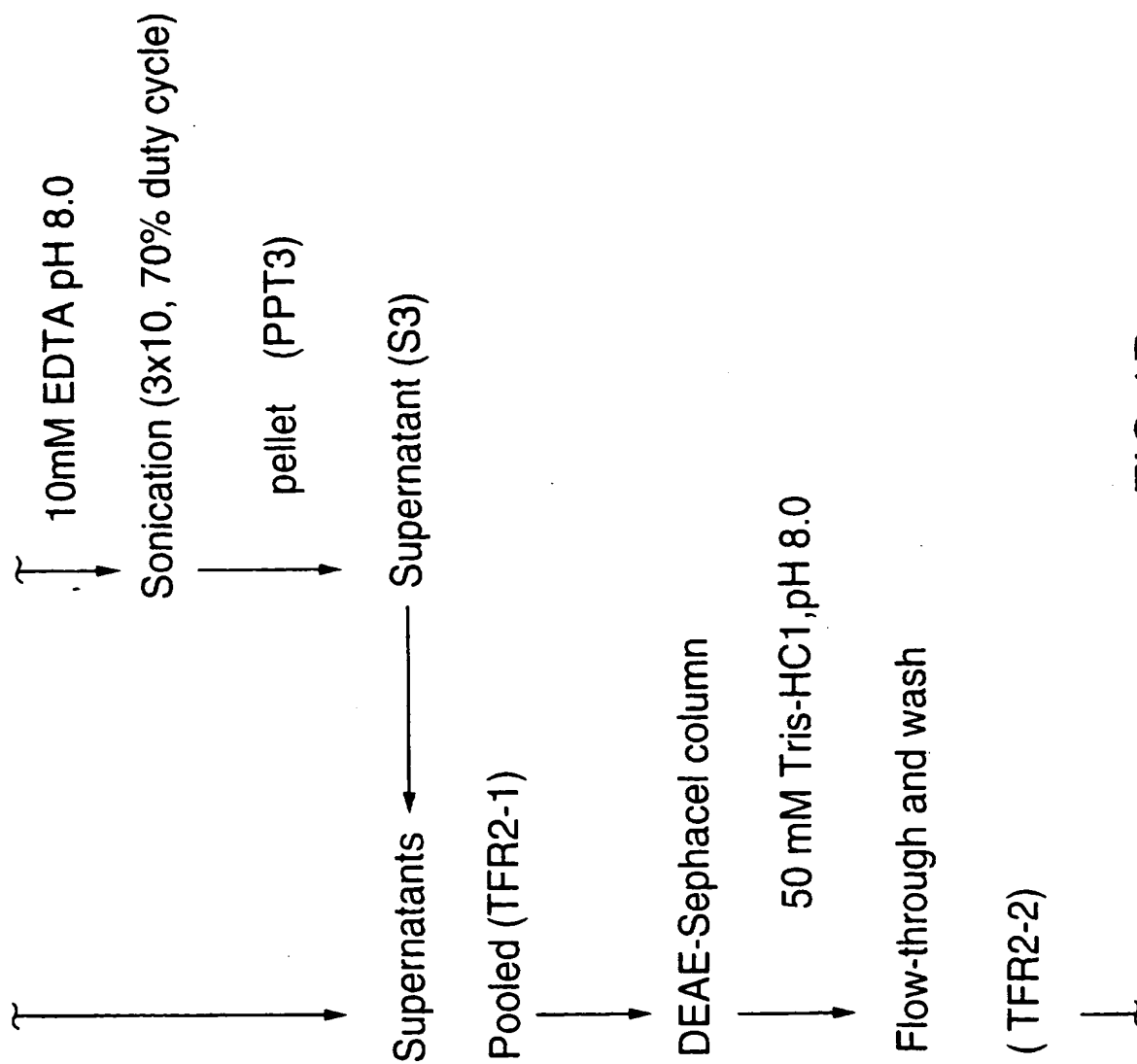


FIG. 1B

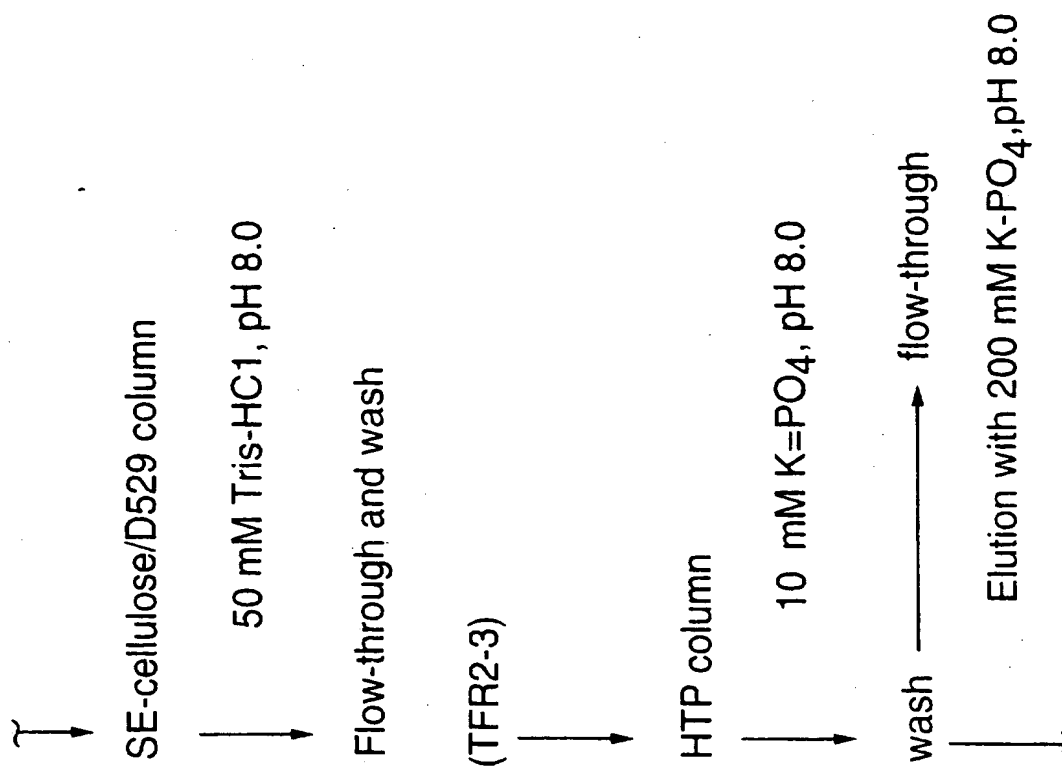


FIG.1C

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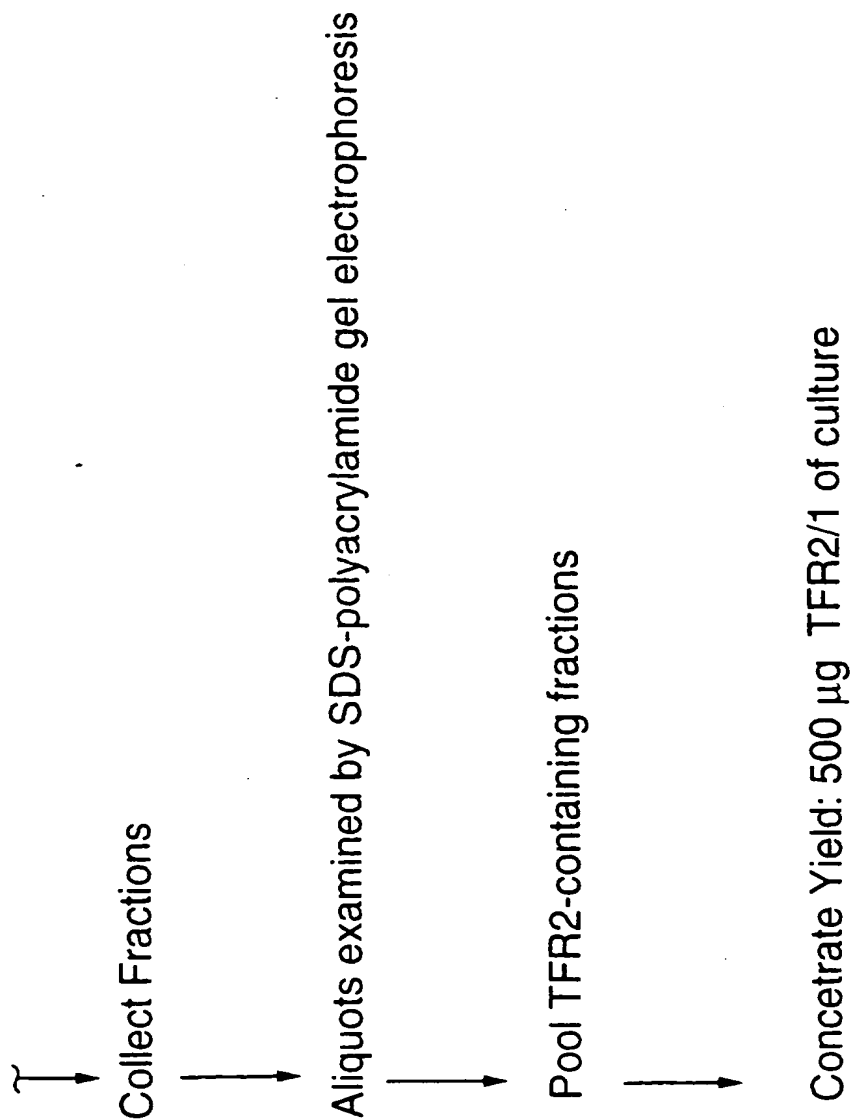
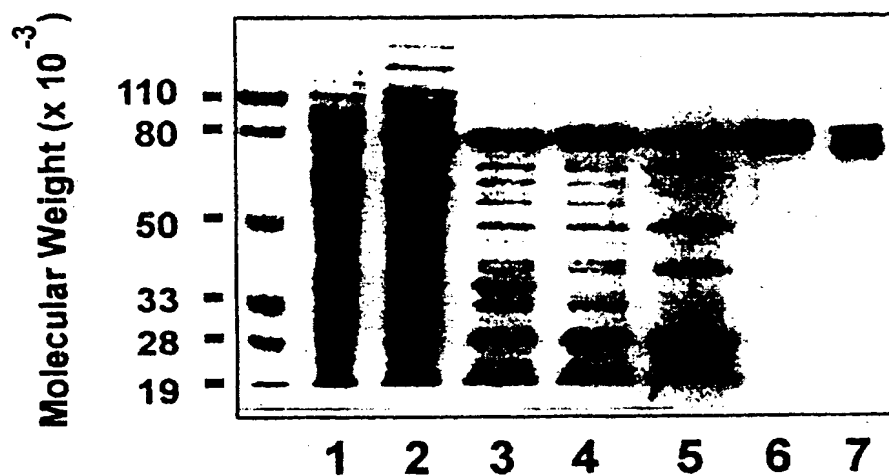


FIG.1D

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1. Lysate of *M. catarrhalis* strain 4223
2. TfR2 extract (TfR2-1)
3. Run-through of DEAE (TfR2-2)
4. Run-through of SE-cellulose/ D529 (TfR2-3)
5. Run-through of HTP (Protein B)
6. HTP bound fraction (TfR2)
7. Affinity purified TfR2

FIG.2.

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FIG.3A.

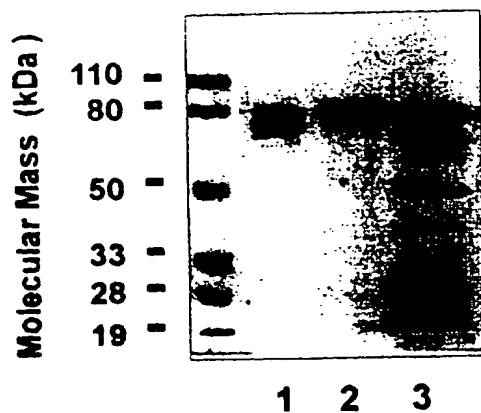
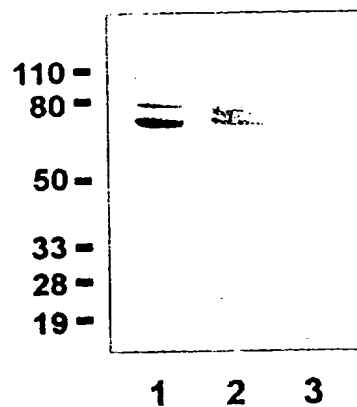


FIG.3B.



1. Affinity purified TfR2
2. TfR2 preparation
3. Protein B preparation

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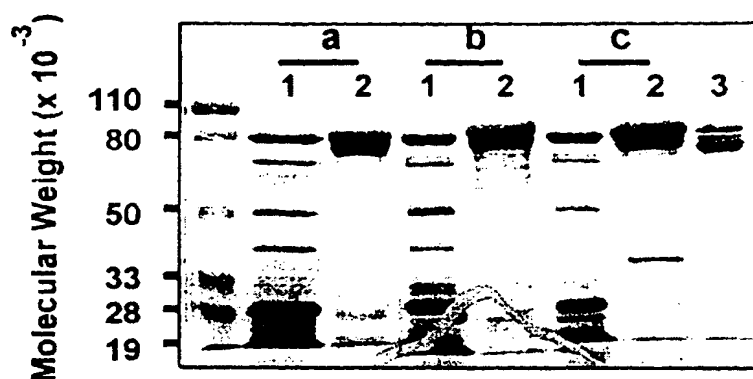


FIG. 4A.

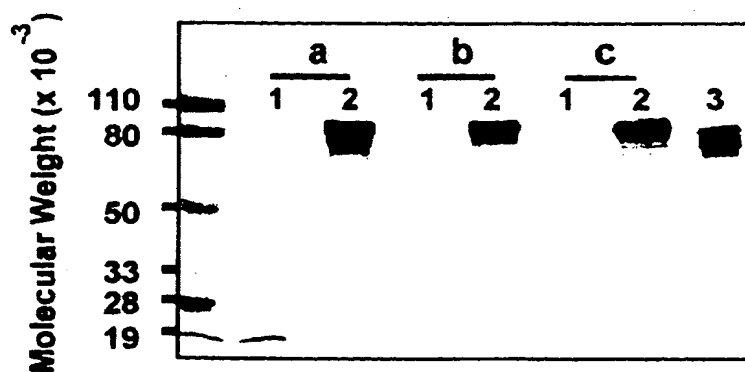


FIG. 4B.

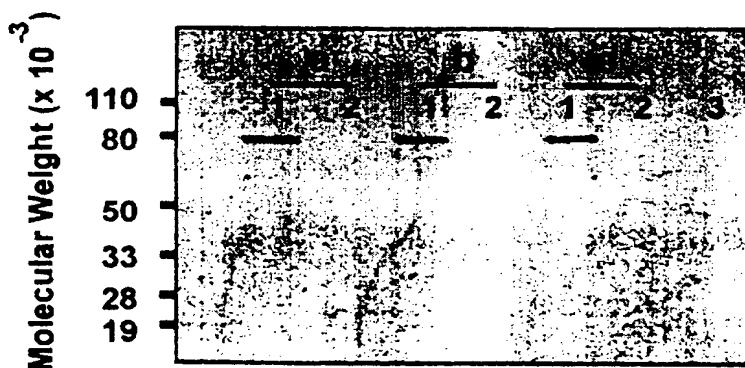


FIG. 4C.

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FIG.5A.

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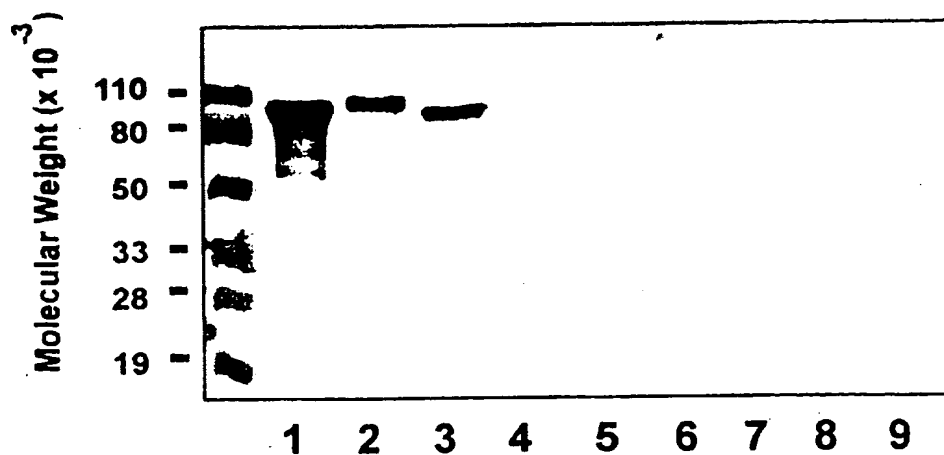


FIG.5B.

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INTERNATIONAL SEARCH REPORT

Internat Application No PCT/LA 96/00684		
A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C07K14/22 C07K16/12 A61K39/095 A61K39/116 A61K39/39 G01N33/569 A61K47/48 //A61K9/16,A61K9/127,A61K9/00,A61K9/48		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 6 C07K A61K G01N		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	MICROBIOL. PATHOGENESIS, vol. 15, 1993, pages 433-445, XP000612196 YU R-H. ET AL.: "The interaction between human transferrin and transferrin binding protein 2 from Moraxella (Branhamella) catarrhalis differs from that of other human pathogens" cited in the application see the whole document --- -/--	1,2,9, 10, 14-21, 26,27, 29-36
<div style="display: flex; justify-content: space-between;"> <div> <input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. </div> <div> <input checked="" type="checkbox"/> Patent family members are listed in annex. </div> </div>		
<div style="display: flex;"> <div style="flex: 1;"> <p>* Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="flex: 1;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
Date of the actual completion of the international search <div style="text-align: center; font-size: 1.2em;">12 December 1996</div>		Date of mailing of the international search report <div style="text-align: center; font-size: 1.2em;">07. 01. 97</div>
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax (+ 31-70) 340-3016		Authorized officer <div style="text-align: center; font-size: 1.2em;">Gac, G</div>

INTERNATIONAL SEARCH REPORT

Application No
PCT/CA 96/00684

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	CAN. J. MICROBIOL., vol. 35, no. 5, May 1989, pages 409-415, XP002020995 SCHRYVERS ET AL.: "Comparative analysis of the transferrin and lactoferrin binding proteins in the family Neisseriaceae" cited in the application see the whole document ---	1,2,9, 10, 14-21, 26,27, 29-36
Y	WO 90 12591 A (UNIVERSITY TECHNOLOGIES INTERNATIONAL INC.) 1 November 1990 see page 8, line 12 - line 35 see page 9 see page 10, line 1 - line 7 see page 11, line 22 - line 36 see page 12 see page 22 - page 26; examples 2,3 ---	1,2,9, 10, 14-21, 26,27, 29-36
A	WO 90 03184 A (MOREIN) 5 April 1990 see the whole document ---	12-18
A	WO 91 06287 A (ENZYTECH INC.) 16 May 1991 see the whole document -----	12,13, 15,16

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA 96/ 00684

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Please see Further Information sheet enclosed.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

Remark: Although claims 20, 21 are directed to a method of treatment of (diagnostic method practised on) the human/animal body (Article 17.2.a)i) and Rule 39.2.iv) PCT) the search has been carried and based on the alleged effects of the compound/composition.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internat Application No

PCT/CA 96/00684

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9012591	01-11-90	US-A- 5292869	08-03-94
		AU-B- 649950	09-06-94
		AU-A- 5526190	16-11-90
		EP-A- 0528787	03-03-93
		JP-T- 4506794	26-11-92
		NZ-A- 247967	24-02-95
		US-A- 5141743	25-08-92

WO-A-9003184	05-04-90	AT-T- 109662	15-08-94
		AU-B- 632067	17-12-92
		AU-A- 4337489	18-04-90
		DE-D- 68917474	15-09-94
		DE-T- 68917474	16-03-95
		EP-A- 0436620	17-07-91
		IE-B- 65428	01-11-95
		JP-B- 2501650	29-05-96
		JP-T- 4501714	26-03-92
		PT-B- 91856	03-07-95

WO-A-9106287	16-05-91	AU-B- 642932	04-11-93
		AU-A- 6736090	31-05-91
		CA-A- 2071867	07-05-91
		EP-A- 0550436	14-07-93
		JP-T- 5500961	25-02-93



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C07K 14/22, 16/12, A61K 39/095, 39/116, 39/39, G01N 33/569, A61K 47/48 // 9/16, 9/127, 9/00, 9/48	A1	(11) International Publication Number: WO 97/13785 (43) International Publication Date: 17 April 1997 (17.04.97)
(21) International Application Number: PCT/CA96/00684 (22) International Filing Date: 11 October 1996 (11.10.96) (30) Priority Data: 08/540,753 11 October 1995 (11.10.95) US (60) Parent Application or Grant (63) Related by Continuation US 08/540,753 (CON) Filed on 11 October 1995 (11.10.95) (71) Applicant (for all designated States except US): CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Avenue West, North York, Ontario M2R 3T4 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): YANG, Yan-Ping [CA/CA]; Apartment 709, 120 Torresdale Avenue, Willowdale, Ontario M2R 3N7 (CA). MYERS, Lisa, E. [CA/CA]; 187 Elizabeth Street, Guelph, Ontario N1E 2X5 (CA). HARKNESS, Robin, E. [CA/CA]; Apartment #1706, 640 Sheppard Avenue East, Willowdale, Ontario M2K 1B8	(CA). KLEIN, Michel, H. [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA). (74) Agent: STEWART, Michael, I.; Sim & McBurney, 6th floor, 330 University Avenue, Toronto, Ontario M5G 1R7 (US). (81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published With international search report.	
(54) Title: TRANSFERRIN RECEPTOR PROTEIN OF MORAXELLA (57) Abstract An isolated and purified non-denatured transferrin receptor protein of a <i>Moraxella</i> strain, particularly <i>M. catarrhalis</i> , has an apparent molecular mass of about 80 to about 90 kDa, as determined by SDS-PAGE. The transferrin receptor protein or a fragment analog thereof is useful in diagnostic applications and immunogenic compositions, particularly for <i>in vivo</i> administration to a host to confer protection against disease caused by a strain of <i>Moraxella</i> .		

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